



(11) Publication number:

0 092 918
A2

(12)

EUROPEAN PATENT APPLICATION

(21) Application number: 83301856.7

(51) Int. Cl.³: **A 61 K 9/22**
A 61 K 47/00

(22) Date of filing: 31.03.83

(30) Priority: 22.04.82 GB 8211704

(43) Date of publication of application:
02.11.83 Bulletin 83/44

(84) Designated Contracting States:
AT BE CH DE FR GB IT LI LU NL SE

(71) Applicant: **IMPERIAL CHEMICAL INDUSTRIES PLC**
Imperial Chemical House Millbank
London SW1P 3JF(GB)

(72) Inventor: **Churchill, Jeffrey Richard**
12 Chiltern Close
Sandiway Northwich Cheshire(GB)

(72) Inventor: **Hutchinson, Francis Gowland**
29 Woodland Drive
Lymm Cheshire(GB)

(74) Representative: **Atkinson, John David et al,**
Imperial Chemical Industries PLC Legal Department :
Patents Thames House North Millbank
London SW1P 4QG(GB)

(54) Continuous release formulations.

(57) Pharmaceutical compositions comprising a pharmacologically active polypeptide and a pharmacologically or veterinarily acceptable amphipathic, non-cross-linked linear, branched or graft block copolymer, which has a minimum weight average molecular weight of 5,000, in which the hydrophobic component is biodegradable and the hydrophilic component may or may not be biodegradable, the composition being capable of absorbing water to form a hydrogel when placed in an aqueous, physiological-type environment; copolymers suitable for use in said compositions; and method for the manufacture of such copolymers.

TITLE: CONTINUOUS RELEASE FORMULATIONS

5 This invention relates to pharmaceutical compositions of pharmacologically-active polypeptides, which provide continuous release of the polypeptide over an extended period when the composition is placed in an aqueous, physiological-type environment.

10 It has long been appreciated that the continuous release of certain drugs over an extended period following a single administration could have significant practical advantages in clinical practice, and compositions have already been developed to provide extended release of a number of clinically useful drugs, after oral dosing (see, for example, Remington's Pharmaceutical Sciences, published by Mack Publishing Company, Easton, 15 Pennsylvania, U.S.A., 15th Edition, 1975, pages 1618-1631), after parenteral administration (*ibidem*, pages 1631-1643), and after topical administration (see, for example, United Kingdom Patent Number 1,351,409). A suitable method of parenteral administration is the sub- 20 dermal injection or implantation of a solid body, for example a pellet or a film, containing the drug, and a variety of such implantable devices have been described. In particular, it is known that, for many drugs, suitable implantable devices for providing extended drug release 25 may be obtained by encapsulating the drug in a biodegradable polymer, or by dispersing the drug in a matrix of such a polymer, so that the drug is released as the degradation of the polymer matrix proceeds.

Suitable biodegradable polymers for use in such sub-
stained release formulations are well known, and
include polyesters which gradually become degraded by
hydrolysis when placed in an aqueous, physiological-type
5 environment. Particular polyesters which have been used
are those derived from hydroxycarboxylic acids, and much
prior art has been directed to polymers derived from α -
hydroxycarboxylic acids, especially lactic acids in both
its racemic and optically active forms, and glycolic acid,
10 and copolymers thereof - see, for example, United States
Patents Numbers 3,773,919 and 3,887,699; Jackanicz et al.,
Contraception, 1973, 8, 227-234; Anderson et al., ibidem,
1976, 11, 375-384; Wise et al., Life Sciences, 1976, 19,
867-874; Woodland et al., Journal of Medicinal Chemistry,
15 1973, 16, 897-901; Yolles et al., Bulletin of the
Parenteral Drug Association, 1976, 30, 306-312; Wise et
al., Journal of Pharmacy and Pharmacology, 1978, 30, 686-
689 and 1979, 31, 201-204.

United Kingdom Patent Specification Number
20 1,325,209 (equivalent to United States Patent
Specification Number 3,773,919) and United States Patent
Specification Number 3,887,669 make reference to extended
or sustained release of polypeptides. The latter
mentions insulin only, but it contains no specific example
25 of any such formulation, and the reference to polypeptides
is apparently entirely speculative, appearing, as it does,
only in an extensive listing of very many different
classes of drugs which can allegedly be incorporated into
formulations of the kind described therein. In fact,
30 essentially all of the other drug types referred to in
that specification, apart from polypeptides, are
relatively hydrophobic in character and of relatively low
molecular weight, and the disclosure of that specification
displays no recognition of the difficulties which we have

encountered when seeking to obtain satisfactory sustained release formulations of polypeptides, many of which are relatively hydrophilic, and of relatively high molecular weight.

5 It is to be appreciated that "sustained" or
"extended" release of a drug may be either continuous or
discontinuous. We have now discovered, in fact, that in
many cases when the teaching of the prior art, and in
particular the teaching of United Kingdom Specification
10 No. 1,325,209, is applied to the manufacture of a
formulation of a polypeptide, the release of the poly-
peptide from the formulation, although occurring over an
extended period of time, may also be discontinuous. For
example, the release of a polypeptide from a polylactide
15 polymer as described in the said Specification is often
preceded by a significant induction period, during which
no polypeptide is released, or is polyphasic, and
comprises an initial period during which some polypeptide
is released, a second period during which little or no
20 poly-peptide is released, and a third period during which
most of the remainder of the polypeptide is released. By
contrast, it is an object of the present invention to
provide compositions of polypeptides from which, apart
possibly from a relatively short initial induction period,
25 the polypeptide is released continuously, with no periods
during which little or no polypeptide is released. The
words "continuous release" are used in this specification
solely to describe a release profile which is essentially
monophasic, although it may have a point of inflection,
30 but certainly has no "plateau" phase.

United Kingdom Patent Specification Number
1,388,580 describes sustained release formulations
containing insulin, which are based on hydrogels formed by

reacting a water soluble polymer with a chelating agent, then cross linking the polymer-chelating agent chains by reaction with a polyvalent metal ion in aqueous solution to form a hydrogel. Insulin was incorporated in a preformed hydrogel in aqueous solution, and the whole was homogenised and injected sub-cutaneously or intramuscularly.

It is an object of the present invention to provide an implantable or injectable pharmaceutical or veterinary formulation for pharmacologically useful polypeptides, which is in solid form, and which absorbs water from the animal body, after implantation, to form a hydrogel from which the polypeptide is released continuously over an extended period of time.

Thus, according to the present invention, there is provided a pharmaceutical composition comprising a pharmacologically useful polypeptide and a pharmaceutically or veterinarily acceptable amphipathic, non-cross-linked linear, branched or graft block copolymer, which has a minimum weight average molecular weight of 5,000, in which the hydrophobic component is biodegradable or hydrolytically unstable under normal physiological conditions, and the hydrophilic component may or may not be biodegradable, the composition being capable of absorbing water to form a hydrogel when placed in water or an aqueous physiological type environment.

This invention is applicable to polypeptides quite generally, without any limitation as to structure or molecular weight, but is most useful for polypeptides which are relatively hydrophilic, and the following list, which is not intended to be exhaustive, is indicative of polypeptides which may be employed in the formulation of

this invention:-

oxytocin, vasopressin, adrenocorticotrophic hormone
(ACTH), epidermal growth factor (EGF), prolactin,
luliberin or luteinizing hormone releasing hormone (LH-
5 RH), growth hormone, growth hormone releasing factor,
insulin, somatostatin, glucagon, interferon, gastrin,
tetragastrin, pentagastrin, urogastrone, secretin,
calcitonin, enkephalins, endorphins, angiotensins, renin,
bradykinin, bacitracins, polymyxins, colistins, tyrocidin,
0 gramicidines, and synthetic analogues and modifications
and pharmaceutically-active fragments thereof, monoclonal
antibodies and soluble vaccines.

A particular LH-RH analogue to which this
invention is applicable is ICI.118,630, [Glu-His-Trp-Ser-
5 Tyr-D-Ser(O-tBu)-Leu-Arg-Pro-Azgly-NH₂].

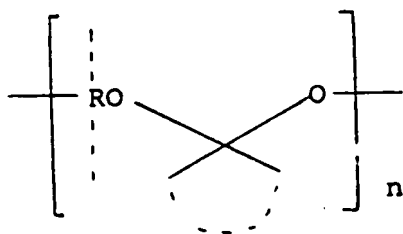
By "an aqueous physiological type environment"
we mean the body, particularly the musculature or the
circulatory system, of a warm-blooded animal, although in
laboratory investigations such an environment may be
0 mimicked by aqueous liquids, optionally buffered to a
physiological pH, at a temperature of between 35 and
40°C.

The continuous release composition of the
invention may be placed in the body of an animal which it
15 is desired to treat with a polypeptide by, for example,
intramuscular or subcutaneous injection, or by sub-dermal
surgical implantation, in conventional clinical or
veterinary manner.

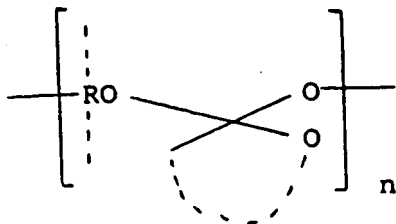
The pharmaceutically or veterinarily acceptable
10 amphipathic copolymer may be, for example, a linear block

copolymer of the formula $A_m(BA)_n$ or $B_m(AB)_n$ wherein m is 0 or 1, n is an integer, A is a pharmaceutically or veterinarily acceptable hydrophobic polymer and B is a pharmaceutically or veterinarily acceptable hydrophilic polymer, or the amphipathic copolymer may be a graft or branched block copolymer of the formula AB_n or BA_n wherein A , B and n have the meanings stated above, and wherein respectively either A or B is a backbone polymer with n units of a monomer or polymer B or A respectively grafted onto it.

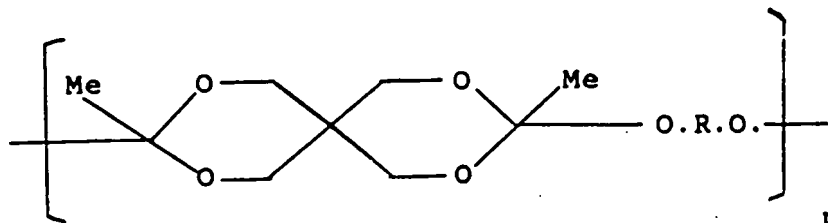
The pharmaceutically or veterinarily acceptable hydrophobic polymer A may be, for example, poly-(D-, L- or DL-lactic acid), poly(D-, L- or DL-lactide), polyglycolic acid, polyglycolide, poly- ϵ -caprolactone, poly(3-hydroxybutyric acid) or a non-therapeutic hydrophobic polypeptide, for example polybenzylglutamate. Alternatively, the hydrophobic polymer A may be a polyacetal of the general formula:



wherein R is a hydrocarbon radical, or a polycarbonate or polyorthoester of the general formula:



wherein R is a hydrocarbon radical, as described in United States Patent Number 4,093,709, which is incorporated herein by reference, or it may be a copolymer comprising such acetal, carbonate, or ortho-ester units alternating
5 with diol units, or it may be a copolymer of the formula:



which is obtained by reacting pentaerythritol with ketene to form 3,9-bis(methylene)-2,4,8,10-tetraoxaspiro[5,5]-undecane which is then copolymerised with a diol
10 HO-R-OH, as described in Journal of Polymer Science, Polymer Letters, 1980, pages 619-624. The diol HO-R-OH may be, for example, a high molecular weight polyethylene glycol or a mixture of that with low molecular weight species, giving a random structure.
15 The hydrophobic polymer may also itself be a copolymer derived from two or more monomers from which the above polymers are derived.

The pharmaceutically or veterinarily acceptable hydrophilic polymer B may be, for example, polyvinyl
20 alcohol, polyvinylpyrrolidone, polyethylene oxide, polyethylene glycol, polyacrylamide, polymethacrylamide, dextran, alginic acid, sodium alginate, gelatine or a copolymer of two or more of the monomers from which the above polymers are derived.

25 In a further alternative, the hydrophilic polymer B may itself be a copolymer, for example a poly-

oxyethylene/polyoxypropylene block copolymer of the type known as "Pluronic" (trade mark) or "Synperonics" (trade mark).

5 Various mechanisms by which drugs are released from biocrodible polymers are described in "Controlled Release of Biactive Materials", edited by R. Baker, Academic Press 1980, particularly chapter 1, pages 1-17, by J. Heller and R.W. Baker.

10 In the present invention, when the dry amphipathic copolymer, containing a polypeptide, is immersed in water or placed in an aqueous physiological environment within an animal body, water uptake is a function of the hydrophilic or water-interactive parts of the copolymer, and the material swells. This absorption
15 of water, however, renders the water-insoluble parts of the copolymer incompatible, and these hydrophobic parts of the copolymer then serve as cross-linking points, which thus serve to limit further water uptake. In this swollen, hydrated state, the matrix is permeable to water-
20 soluble polypeptides incorporated within the matrix, and such polypeptides are thus progressively desorbed from the matrix.

In the process of swelling, and when swollen to some equilibrium state, hydrolytic degradation of the
25 hydrophobic part of the copolymer starts to occur. The partially degraded copolymer has greater swellability, so that continued hydrolysis leads to progressively further water uptake, an increasingly water-permeable matrix, and a further increase in polypeptide desorption which
30 compensates for its decreasing concentration and maintains its continuous release. Thus, by appropriate design of the copolymer material, the initial swelling to a hydrogel

and consequent desorption of active material, and the rate of subsequent hydrolytic degradation to increase the further desorption of active material to compensation for its decreasing concentration in the matrix, can be controlled so as to give continuous release of active material over an extended period of time, as defined above.

Such an ideal release profile for the active material can also be obtained by blending different copolymers, each having its own defined properties (for example, molecular weight, molecular weight distribution, block structure, hydrophilicity, degradation properties, diffusional properties), and by appropriate combination of different such materials, the rate of release, and the duration of release, of an active material can be varied as desired.

Also by appropriate choice of the above parameters, and/or appropriate blending, a copolymer material can be obtained which allows of processing into implants at relatively low temperatures, certainly below 100°C., and in some cases even at room temperature, and is thus suitable for the fabrication of implants incorporating heat sensitive or solvent sensitive polypeptide active materials. For example, block copolymers of polyethylene glycol and amorphous hydrophobic polymers, having a glass transition temperature above 37°C., are particularly useful, because the polyethylene glycol block plasticises the hydrophobic block, giving a material which is readily processed at relatively low, even at room temperature, while on subsequent standing the polyethylene glycol blocks crystallise to give a tough hard product which can be easily handled.

The block copolymers defined above are themselves novel, useful materials, per se. Thus, according to a further feature of the invention, there is provided a pharmaceutically or veterinarily acceptable
5 amphipathic linear, branched or graft block copolymer, which has a minimum weight average molecular weight of 5,000, in which the hydrophobic component is biodegradable or hydrolytically unstable under normal physiological conditions, and the hydrophilic component may or may not
10 be biodegradable, and which copolymer is capable of absorbing water to form a hydrogel when placed in water on an aqueous environment.

Particular such copolymers are those defined above.

15 According to a further feature of the invention, there is provided a blend of two or more such copolymers as defined above.

These copolymers and copolymer blends are also useful more generally for the continuous release of non-
20 peptide drugs by oral, including intra-ruminal, parenteral, ocular, rectal or vaginal administration.

Thus, according to further features of this invention there are provided pharmaceutical or veterinary compositions comprising a non-peptide pharmacologically-
25 active compound and a block copolymer as defined above, and the use of such block copolymers for the continuous release of such a non-peptide, pharmacologically-active compound.

According to a further feature of the invention, there is provided a process for the manufacture of a pharmaceutically or veterinarily acceptable amphipathic linear, branched or graft block copolymer as defined above, which comprises copolymerising monomer A and monomer B by conventional techniques such as graft copolymerisation, polycondensation and polyaddition, optionally with an appropriate catalyst, for example zinc oxide, zinc carbonate, basic zinc carbonate, diethyl zinc, organotin compounds, for example stannous octoate (stannous 2-ethylhexanoate), tributylaluminium, titanium, magnesium or barium compounds or litharge, and of these stannous octoate is generally preferred.

The copolymerisations are otherwise carried out in conventional manner, known in the polymer art, as regards time and temperature.

The invention is illustrated but not limited by the following Examples:-

Example 1

Polyethylene glycol of molecular weight 20,000 (30g.) was stirred and heated under vacuum (<0.1 mm. of mercury) at 120°C . for 3 hours. D,L-lactide (15g.) and glycolide (15g.) were added, and the mixture was stirred under a nitrogen atmosphere until the solids melted. The temperature was raised to 160°C ., and stannous octoate (stannous 2-ethylhexanoate) (0.1ml.) was added. The mixture was maintained at 160°C . for 3 hours, by which time it had become highly viscous, and it was then cooled and dissolved in acetone (200ml.). This acetone solution was added slowly to vigorously stirred ethanol (1500ml.), and the precipitate thus produced was filtered off, and

dried in a vacuum oven for 3 hours at room temperature, then overnight at 40°C.

The n.m.r. spectrum of this copolymer, in deuteriochloroform, showed it to have the composition oxyethylene:lactic:glycolic of 2:1:1.

This copolymer was moulded at about 60°C. to a soft, plastic transparent film. A sample (39mg.) swelled rapidly when placed in water, taking up 135mg. of water over 4 hours, to give a transparent hydrogel which subsequently disintegrated over 2 weeks at 37°C.

Example 2

The polymer described in Example 1 (20.2mg.) and bovine growth hormone (BGH), (5.1mg.) were blended together at about 40°C. to give an opaque blend, which was moulded into a slab 1mm. thick. This slab was immersed in a buffer solution at pH 8.6 (M/15 buffer, pH 8.6, containing 0.01% sodium azide), and released a material of molecular weight about 22,000 which had the same retention time on high pressure liquid chromatography as BGH, over a period of at least 12 days.

Example 3

Using the process described in Example 2, the copolymer/BGH blend was formed into discs weighing about 45mg. and containing about 20% of BGH. When such discs were implanted, each in a hypophysectomised rat, the animals' weights increased by an average of 25% over 7 days, whereas the weights of control animals each given a placebo implant remained virtually unchanged.

Example 4

The copolymer described in Example 1 (13.5mg.) and monoclonal mouse immunoglobulin A, (IgA), with a defined antigen specificity and a molecular weight of ≥ 180000 (1.5mg.) were blended at 50°C. to give a homogeneous mixture of the IgA in the copolymer, and the protein/copolymer mixture was moulded to give a sphere of ~ 2 mm. diameter. The in vitro release of IgA was evaluated by immersing the protein copolymer in buffer (phosphate buffered saline, pH 7.2) at 37°C. Using an enzyme linked immunoassay technique the aqueous medium was assayed for active IgA, and release of the biologically active protein was shown to start after 2 days and continue for at least 9 days.

15 Example 5

Polyethylene glycol having a molecular weight of 20000 (50g.) was dissolved in chloroform (150mg.) and washed six times with distilled water (~ 300 ml.), discarding the aqueous washes. The chloroform was evaporated under reduced pressure and the purified polyethylene glycol was dried at 160°C/0.05mm.Hg. for 1hr.

Stannous octoate (stannous 2-ethylhexanoate) (~ 5 g.) was purified by heating at 140°C/0.055mm.Hg. to remove impurities. The purified polyethylene glycol (14.3g.) was heated to 160°C. under vacuum (0.05mm.Hg.) in a 100ml. round bottomed flask for 1 hr. Freshly prepared, pure D,L-lactide (42.9g.) was added under nitrogen and melted at 160°C. Stannous octoate (0.2ml.) was added, and the mixture was stirred until the viscosity no longer allowed stirring to continue. After

3 hrs., a highly viscous product was obtained. The mixture was allowed to cool, the flask was broken, the contents of the flask were dissolved in acetone (~300ml.) and the solution was filtered. The filtrate was added slowly to ethanol (~1000ml.) with vigorous agitation to give a fibrous precipitate, which was collected and dried at 30°C. in a vacuum oven overnight. Analysis of the product by n.m.r. showed the product to have a composition of oxyethylene:lactic of 1:3, and the intrinsic viscosity in chloroform was 1.055.

The product was moulded to a thin (0.2mm.), soft, plastic transparent film. On immersion in water, the film (0.54g.) increased in weight to 0.95g. in 1 day at 37°C. The hydrated transparent film had rigidity and strength superior to the initially dry copolymer. After 35 days, the film was intact and retained good mechanical properties showing that the copolymer was being degraded only slowly, as shown by change of composition by n.m.r.

When bovine growth hormone is incorporated into the dry copolymer at 60° C., the resulting polypeptide/polymer blend releases a 22000 molecular weight product into buffer (M/15 phosphate buffer, pH 8.6) over at least 7 days.

25 Example 6

Polyethylene glycol having a molecular weight of 6000 (50g.) was purified using the method described above in Example 5.

The purified dry polyethylene glycol (7.5g.) and stannous chloride dihydrate (15mg.) were mixed at room temperature then heated with stirring to 155°C. under high vacuum (0.1-0.01mm.Hg.) and maintained at this temperature for 2 hrs. while freshly prepared, dry D,L-lactide (22.5g.) was added to the mixture under nitrogen, and melted. The reaction temperature was maintained at 155-160°C. for 3 hrs. to give a viscous product, which was poured onto poly-tetrafluoroethylene film and allowed to cool. The polymeric product was dissolved in acetone (70ml.) with warming, and the polymer was isolated by pouring the acetone solution into ethanol (600ml.). The precipitate was dried in a vacuum oven overnight at 60°C. The polymer had an intrinsic viscosity in chloroform of 0.41. When pressed as a thin film (0.2mm.) and immersed in water, the polymer takes up approximately its own weight of water at 37°C. over 24 hrs. to give a tough hydrogel.

Example 7

99Mg. of a block copolymer as prepared in Example 6 containing 25 parts of polyethylene glycol (mol. wt. 6000) and 75 parts of poly(D,L-lactide) was dissolved in 4.5ml. of anhydride free glacial acetic acid and 0.5ml. of distilled water. 200 μ L. of a solution containing 1.1mg. of mouse epidermal growth factor (EGF) was added to the polymer solution, and the mixture was homogenised. The homogenised solution was frozen and then freeze dried for 18 hrs., the product was moulded at 50°C. to give an implant weighing 40mg. (~ 8mm. x 4mm. x 1mm). The implant was placed in 1ml. of human serum at 37°C., and the release of EGF was measured by radio immunoassay on aliquots of serum. The results showed a continuous release of peptide over at least three days.

Example 8

25G. of a copolymer containing equimolar proportions of D,L-lactide and glycolide and having an intrinsic viscosity in chloroform of 0.20 was dissolved in 50ml. of dry ethyl acetate, and the solution was heated to reflux with stirring under nitrogen. 0.25G. of lauroyl peroxide was dissolved in freshly distilled vinyl pyrrolidone (25ml.). The mixture was added dropwise to the refluxing polymer over 2 hrs., and the mixture was heated at reflux for a further 6 hrs. On cooling, the mixture gelled. Purification of the amphipathic block graft copolymer by removal of homo polymer of polyvinyl pyrrolidone using precipitation techniques was difficult as precipitation often resulted in colloidal suspensions, and this indicated that grafting of polyvinyl pyrrolidone to the lactide/glycolide copolymer had occurred.

The ethyl acetate mixture was therefore warmed to 70°C. and 50ml. of ethanol was added to give a colloidal suspension, from which the polymer was isolated by precipitation into n-hexane (2 litres). The polymer thus obtained was dried at 90°C. overnight under vacuum to give on cooling a brittle product consisting of graft copolymer and homo polyvinylpyrrolidone. The product had an intrinsic viscosity of 0.29 in chloroform, and approximately 50% of polyvinyl-pyrrolidone as homo-copolymer and graft block copolymer.

The polymer thus obtained (0.45g.) and ICI. 118,630 (0.05g.) were dissolved in anhydride-free glacial acetic acid (5ml.) and freeze dried at 0.01mm. of mercury for 22 hr.

The product was moulded at 110°C. for 20secs. to give a slab (~0.8cm. x 1.2mm. x 2mm., weighing 30mg.) which, when immersed in aqueous pH 7.4 buffer at 37°C. released the peptide over a period of several days.

5 Example 9

50G. of polyvinyl alcohol having a molecular weight of 14,000 was dissolved in 500g. of commercial D,L-lactic acid (containing ~12% water) with stirring under nitrogen. The mixture was heated to 140°C. and water
10 was distilled off over 8hrs., during which time the mixture became progressively more viscous and its temperature rose to 190°C. When no further water distilled over, the pressure was reduced to ~25cm. of mercury, and the mixture was heated for a further 8hrs.
15 Finally, the pressure was reduced to 0.1mm. of mercury and the mixture was heated at 200°C. for 8 hrs. to give a highly viscous amber product.

The polymer was allowed to cool, and the flask was broken. The product was broken up into small pieces
20 and dissolved in methanol (1.5 litres), and the product was isolated by precipitation in 10 litres of distilled water. The precipitation was washed with a further 5 litres of water, and dried under vacuum at room temperature for 8 hrs., finally at 100°C. for 16hrs., to
25 give an amber glassy product which consisted of a polyvinyl alcohol back-bone containing pendant polylactic acid chains of low molecular weight, intrinsic viscosity = 0.65 in chloroform. The product contained approximately 85% of polylactic acid, and the pendant polylactic acid
30 average chain length was approximately 3.5.

The polymer was moulded at 100°C. to give a slab 1cm. x 0.2cm. x 0.2cm. which was immersed in water at 37°C. The product absorbed water and became flexible, and eroded to give soluble products over a period of 2 months.

Example 10

Mouse epidermal growth factor (285 μ l. of a 21mg./ml. solution in distilled water) was added to a solution of an 80/20 poly(D,L-lactide)/PEG 6000 copolymer of intrinsic viscosity 0.36 in chloroform, (45mg.) in 2.5ml. of 90% aqueous acetic acid. The solution of peptide and polymer was frozen and then freeze dried at ~ 0.01mm.Hg. for 24 hours to give a dried product. The freeze dried material was moulded at 60°C. to give implants weighing 13.5mg. and 15.3mg. containing 1.3 and 1.5mg. of peptide respectively.

These were implanted subcutaneously into 2 cats, each fitted with a gastric fistula. Blood samples were taken, and gastric acid output in response to a histamine stimulus was measured. Peptide was detected in the blood by radioimmunoassay for a minimum of 3 days subsequent to implantation, and gastric acid output showed inhibition of 3-6 days subsequent to implantation.

Example 11

Mouse epidermal growth factor (120 μ l. of 10.5mg. peptide in 320 l. of distilled water) was added to a solution of an 85/15 poly(D,L-lactide)/PEG 6000 copolymer of intrinsic viscosity 0.39 in chloroform, (36mg.) in 1.8ml. of 90% aqueous acetic acid. The resultant solution was frozen and freeze-dried overnight. The

freeze dried material was moulded at 70°C. to give an implant weighing 16.9mg. (dimensions approximately 1 x 1 x 5mm.).

Peptide was released from this implant
5 continuously over at least 15 days into 10% human serum in water containing 0.1% sodium azide.

Example 12

To show the effect of composition and hydro-
philicity of the block copolymer on release of polypeptide
10 from implants the following comparison was carried out.

In separate experiments implants were prepared using

- (a) a block copolymer of intrinsic viscosity 0.39 in chloroform containing 25% w/w polyethylene glycol
15 having a molecular weight of 6000 and 75% w/w of poly(D,L-lactide).
- (b) a block copolymer of intrinsic viscosity 0.79 in chloroform containing 5% w/w polyethylene glycol
20 having a molecular weight of 6000 and 95% w/w of poly(D,L-lactide).

76.2mg. of polymer and ICI 118,630 (23.8mg. as the acetate salt, equivalent to 20mg. pure peptide) were dissolved in anhydride-free glacial acetic acid (1.5ml.). The solution was frozen and freeze dried for 18 hours, and
25 the freeze dried product was moulded at ~70°C. to give implants weighing ~ 45-50mg. (dimensions approximately 0.2cm. x 0.2cm. x 1cm.).

The implants were immersed in 1ml. of McIlvains pH 7.4 buffer at 37°C., and 1ml. samples of the aqueous
30 medium were removed at given time points and assayed by

high pressure liquid chromatography for drug content.
The aqueous medium removed was replaced each time by 1ml.
of fresh buffer.

5 These release experiments showed that implants
prepared using the more hydrophilic is polymer, containing
25% polyethylene glycol, released compounds for ~ 18
days.

10 In contrast, the implant prepared using the less
hydrophilic copolymer, containing 5% polyethylene glycol,
released compound continuously for at least 250 days.

Example 13

Poly(ethylene glycol methyl ether) having a
molecular weight of 5000 was purified as in Example 5.

15 20G. of the purified poly(ethylene glycol methyl
ether) was dried at 160°C./0.01mm.Hg. for 1 hour. 80G.
of dry, freshly prepared D,L-lactide were added and the
mixture stirred under a nitrogen atmosphere at 160°C.
When all the D,L-lactide had melted, 0.15ml. of stannous
octoate (stannous-2-ethylhexanoate) were added, and the
20 mixture was maintained at 160°C. for 6 hours, during
which time a highly viscous product was formed. The
mixture was allowed to cool, the flask was broken and the
contents were dissolved in 200ml. of acetone. The
actone solution of polymer was added with vigorous
25 stirring to 2000ml. of hexane to precipitate the polymer.
The precipitated polymer was dried at 70°C. under
reduced pressure for 24 hours to give a block copolymer
having an AB structure where A is polylactide and B is
poly(ethylene glycol methyl ether).

This copolymer is particularly useful for preparing water-in-oil dispersions, which can be used to prepare microcapsules, or in microencapsulation procedures.

5 For example, 5g. of the copolymer was dissolved in 200ml. of methylene chloride, and 1ml. of an aqueous solution of ICI.118,630 containing 20mg. of compound was added with vigorous stirring, to produce a stable water in oil dispersion.

10 The water-in-oil emulsion was stirred vigorously and a non-solvent, such as hexane (2000ml.) was added slowly to produce microcapsules, which were isolated by filtration, and dried, to give a drug/polymer mixture which even in this microcapsule or microencapsulated form
15 gives sustained release over a period of several days.

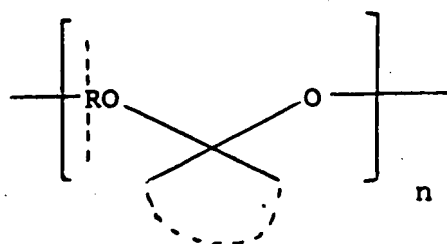
 The poly(ethylene glycol methyl ether) used in the above process was replaced by other derivatives of poly(ethylene glycol) to prepare similar block copolymers, and suitable examples are the monocetyl ethers (ceto
20 macrogols) and stearate esters.

What we claim is:-

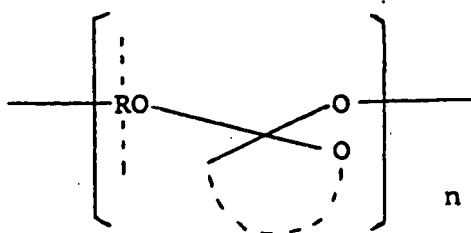
1. A pharmaceutical composition comprising a pharmacologically useful polypeptide and a pharmaceutically or veterinarily acceptable, amphipathic, non-cross-linked linear, branched or graft block copolymer, which has a minimum weight average molecular weight of 5,000, in which the hydrophobic component is biodegradable or hydrolytically unstable under normal physiological conditions, and the hydrophilic component may or may not be biodegradable, the composition being capable of absorbing water to form a hydrogel when placed in water or an aqueous physiological-type environment in an animal body.
2. A composition as claimed in claim 1 wherein the pharmacologically useful polypeptide is selected from oxytocin, vasopressin, adrenocorticotrophic hormone, epidermal growth factor, prolactin, luliberin or luteinising hormone releasing hormone, growth hormone, growth hormone releasing factor, insulin, somatostatin, glucagon, interferon, gastrin, tetragastrin, pentagastrin, urogastrone, secretin, calcitonin, enkephalins, endorphins, angiotensins, renin, bradykinin, bacitracins, polymyxins, colistins, tyrocidin, gramicidines, and synthetic analogues, modifications and pharmacologically active fragments thereof, monoclonal antibodies and soluble vaccines.
3. A composition as claimed in claim 1 wherein the poly-peptide is \square Glu-His-Trp-Ser-Tyr-D-Ser(O-tBu)-Leu-Arg-Pro-Azgly-NH₂.
4. A composition as claimed in claim 1, 2 or 3 wherein the pharmaceutically or veterinarily acceptable

amphipathic copolymer is a linear block copolymer of the formula $A_m(BA)_n$ or $B_m(AB)_n$ wherein m is 0 or 1, n is an integer, A is a pharmaceutically or veterinarily acceptable hydrophobic polymer and B is a pharmaceutically or veterinarily acceptable hydrophilic polymer, or the amphipathic copolymer is a graft or branched block copolymer of the formula AB_n or BA_n wherein A, B and n have the meanings stated above and wherein respectively either A or B is a backbone polymer with n units of a monomer or polymer B or A respectively grafted onto it.

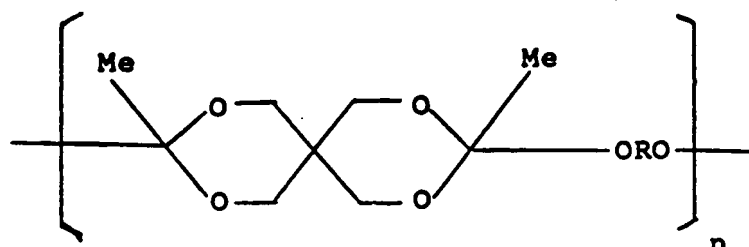
5. A composition as claimed in claim 4 wherein the pharmaceutically or veterinarily acceptable hydrophobic polymer A is selected from poly(D-, L- and DL-lactic acids), poly(D-, L- and DL-lactides), polyglycolic acid, polyglycolide, poly-ε-caprolactone, poly(3-hydroxybutyric acid), non-therapeutic hydrophobic polypeptides, poly-acetals of the formula:-



wherein R is a hydrocarbon radical and n has the meaning stated above, polycarbonates or polyorthoesters of the formula:-



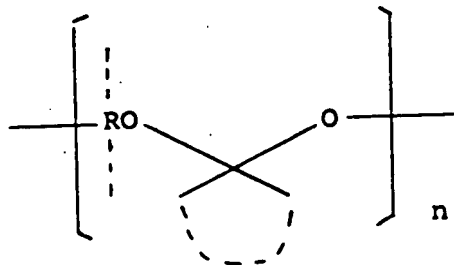
wherein R and n have the meaning stated above, and copolymers of the formula:-



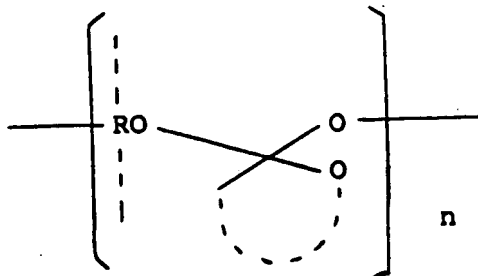
- wherein R and n have the meanings stated above, and
- 5 copolymers derived from two or more monomers from which the above polymers are derived; and the pharmaceutically or veterinarily acceptable hydrophilic polymer B is selected from polyvinyl alcohol, polyvinylpyrrolidone, polyethylene oxide, polyethylene glycol, polyacrylamide, polymethacrylamide, dextran, alginic acid, sodium
- 10 alginate, and gelatine, copolymers of two or more of the monomers from which the above polymers are derived, and polyoxyethylene/polyoxypropylene block copolymers.
6. A pharmaceutically or veterinarily acceptable
- 15 amphipathic, non-cross-linked linear, branched or graft block copolymer, which has a minimum weight average molecular weight of 5,000, in which the hydrophobic component is biodegradable or hydrolytically unstable under normal physiological conditions, and the hydrophilic
- 20 component may or may not be biodegradable, the copolymer being capable of absorbing water to form a hydrogel when placed in water or an aqueous physiological-type environment in an animal body.
7. A copolymer as claimed in claim 6 wherein the
- 25 pharmaceutically or veterinarily acceptable amphipathic copolymer is a linear block copolymer of the formula

$A_m(BA)_n$ or $B_m(AB)_n$ wherein m is 0 or 1, n is an integer, A is a pharmaceutically or veterinarily acceptable hydrophobic polymer and B is a pharmaceutically or veterinarily acceptable hydrophilic polymer, or the amphipathic copolymer is a graft or branched block copolymer of the formula AB_n or BA_n wherein A , B and n have the meanings stated above and wherein respectively either A or B is a backbone polymer with n units of a monomer or polymer B or A respectively grafted onto it.

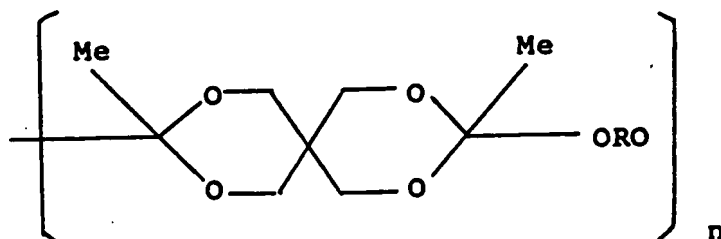
- 10 8. A copolymer as claimed in claim 7 wherein the pharmaceutically or veterinarily acceptable hydrophobic polymer A is selected from poly (D-, L- and DL-lactic acids), poly(D-, L- and DL-lactides), polyglycolic acid, polyglycolide, poly- ϵ -caprolactone, poly(3-hydroxy-
15 butyric acid), non-therapeutic hydrophobic polypeptides, polyacetals of the formula:-



wherein R is a hydrocarbon radical, and n has the meaning stated above, polycarbonates or polyorthoesters of the
20 formula:-



wherein R and n have the meanings stated above, and copolymers of the formula:-



wherein R and n have the meanings stated above, and
 5 copolymers derived from two or more monomers from which
 the above polymers are derived; and the pharmaceutically
 or veterinarily acceptable hydrophilic polymer B is
 selected from polyvinyl alcohol, polyvinylpyrrolidone,
 polyethylene oxide, polyethylene glycol, polyacrylamide,
 10 polymethacrylamide, dextran, alginic acid, sodium
 alginate, and gelatine, and copolymers of two or more of
 the monomers from which the above polymers are derived,
 and polyoxyethylene/polyoxypropylene block copolymers.

9. A process for the manufacture of a copolymer as
 15 claimed in claim 7 which comprises copolymerising monomer
 or polymer A and monomer or polymer B by graft
 copolymerisation, polycondensation or polyaddition.

10. A pharmaceutical or veterinary composition
 comprising a non-peptide, pharmacologically active
 20 compound and a copolymer as claimed in claim 6.

DS32265

SB05

10 Mar 83

John D. Wilkinson
 JOHN D. WILKINSON
 AUTHORIZED REPRESENTATIVE
 General Authorization No. 97